

# Evolutionary development of three gonadotropin-releasing hormone (GnRH) systems in vertebrates

E. A. Dubois, M. A. Zandbergen,\* J. Peute and H. J. Th. Goos

Research Group of Comparative Endocrinology, Graduate School for Developmental Biology, Faculty of Biology, Utrecht, The Netherlands

**ABSTRACT:** Gonadotropin-releasing hormone (GnRH) is the neuropeptide that links the brain to the reproductive system. Most vertebrate species express two forms of GnRH, which differ in amino acid sequence, localization, distribution, and embryological origin. The GnRH system in the ventral forebrain produces a species-specific GnRH form and projects toward the gonadotropic cell in the pituitary. The GnRH neurons of this system originate from the olfactory placode and migrate into the brain during early development. The other GnRH system is localized in a nucleus in the midbrain, where large cells express chicken-GnRH-II, of which the function is still unclear. In modern teleosts, a third GnRH system is present in the terminal nerve, which contains salmon GnRH. The three GnRH systems appear at different times during fish evolution. Besides the two accepted lineages in GnRH evolution (of conserved chicken GnRH-II in the midbrain and of mammalian GnRH or species-specific GnRH in the hypophysiotropic system), we propose a third lineage: of salmon GnRH in the terminal nerve. © 2002 Elsevier Science Inc.

**KEY WORDS:** Gonadotropin-releasing hormone, Evolution, Reproduction, Neuroendocrinology, Brain.

## INTRODUCTION

The decapeptide gonadotropin-releasing hormone (GnRH) forms an essential link in the integration of the external and internal stimuli in the control of reproduction in all vertebrates. The brain integrates the available information and transfers the signal to the GnRH neurons: when to release their GnRH content to allow an adequate response by the reproductive axis. GnRH reaches the gonadotropic cells either indirectly via the portal system (in tetrapods) or directly via axon endings (in most teleosts). In the pituitary, GnRH stimulates the synthesis and release of the gonadotropic hormones FSH (follicle-stimulating hormone) and LH (luteinizing hormone). Besides this endocrine function, GnRH probably fulfills a neuromodulatory role in sexual behavior and in transferring olfactory signals. Because it plays these pivotal roles in all vertebrates, it is well known that GnRH is highly conserved throughout evolution.

## GNRH: FORMS, GENES, AND PHYLOGENY

The first reports about the existence of a GnRH-like substance date from 1971 [1,2,29]. Since this neuropeptide was first identified in mammals (pig and sheep), it is now known as mammalian GnRH (mGnRH). The original name was luteinizing hormone

releasing hormone (LHRH), referring to its stimulatory effect on LH release. We now know that also FSH release is stimulated by the peptide, hence its more general name, GnRH. Until now, 15 forms of GnRH have been identified in various vertebrates and in a protochordate (Table 1). Generally, the GnRHs are named after the species in which they were first discovered. To date, six GnRHs are isolated from fish species (Table 1): salmon GnRH (sGnRH) [52], catfish GnRH (cfGnRH) [3], dogfish GnRH (dfGnRH) [25], seabream GnRH (sbGnRH) [46], herring GnRH (hGnRH) [5], and medaka GnRH (mdGnRH) [36]. Primitive species such as the lamprey and the protochordate *Ciona intestinalis* have their own forms of GnRH: lamprey GnRH I and III [55,57] and tunicate GnRH I and II, respectively [9,44]. Chicken GnRH-I (cGnRH-I) [19] and chicken GnRH-II (cGnRH-II) [31] were both first characterized in chicken. Apart from the common mGnRH, recently the guinea pig GnRH (gpGnRH) was shown as a novel and alternative form of mammalian GnRH [16]. The most recent finding of a novel GnRH was made in an amphibian, the frog *Rana dybowskii*: ranid GnRH (rGnRH) [66].

The GnRH gene consists of three introns and four exons. The second, third, and part of the fourth exon encode for the pre-pro-hormone, which contains a signal peptide (21–23 amino acids), the GnRH (10 amino acids), a cleavage site (Gly-Lys-Arg), and the GnRH-associated peptide (GAP, 40–60 amino acids) [20,21]. The sequence of the second exon is the most conserved, whereas the other exons show high variability. As a consequence, the signal peptides and the GnRHs are well conserved, but the GAPs show less homology among species.

In all classes of vertebrates and in a few invertebrates, the presence of given GnRH forms was established (Table 2). GnRH is a functionally old and well-conserved peptide, because it already controls reproductive functions in molluscs [67] and protochordates [12,20].

In general, all investigated species to date possess two or three different forms of GnRH (Table 2). The most conserved form of GnRH is cGnRH-II, and it coexists in all classes of vertebrates from the *Chondrichthyes* species onward, together with a species-specific GnRH and a possible third form. The two or three forms of GnRH coexisting in one species are transcribed from different genes. The species-specific forms vary, for example, cGnRH-I in birds, hGnRH in herring, dfGnRH in sharks, sGnRH in salmonids, and mGnRH in primates. But, if a third form is present, as shown for “modern” fishes, it is always the sGnRH form.

\* Address for correspondence: Dr. M. A. Zandbergen, Research Group of Comparative Endocrinology, Graduate School for Developmental Biology, Faculty of Biology, Padualaan 8, 3584 CH Utrecht, The Netherlands. Fax: +31 30 2532837; E-mail: M.A.Zandbergen@bio.uu.nl

TABLE 1  
AMINO ACID SEQUENCES OF 15 GnRH FORMS

GnRH	1	2	3	4	5	6	7	8	9	10	
Chicken II	pGlu	His	Trp	Ser	His	Gly	Trp	Tyr	Pro	Gly-NH <sub>2</sub>	[31]
Mammalian	pGlu	His	Trp	Ser	<b>Tyr</b>	Gly	<b>Leu</b>	<b>Arg</b>	Pro	Gly-NH <sub>2</sub>	[2,29]
Rana	pGlu	His	Trp	Ser	<b>Tyr</b>	Gly	<b>Leu</b>	<b>Trp</b>	Pro	Gly-NH <sub>2</sub>	[66]
Catfish	pGlu	His	Trp	Ser	His	Gly	Leu	Asn	Pro	Gly-NH <sub>2</sub>	[3]
Salmon	pGlu	His	Trp	Ser	<b>Tyr</b>	Gly	Trp	<b>Leu</b>	Pro	Gly-NH <sub>2</sub>	[52]
Herring	pGlu	His	Trp	Ser	His	Gly	<b>Leu</b>	<b>Ser</b>	Pro	Gly-NH <sub>2</sub>	[5]
Medaka	pGlu	His	Trp	Ser	<b>Phe</b>	Gly	<b>Leu</b>	<b>Ser</b>	Pro	Gly-NH <sub>2</sub>	[36]
Seabream	pGlu	His	Trp	Ser	<b>Tyr</b>	Gly	<b>Leu</b>	<b>Ser</b>	Pro	Gly-NH <sub>2</sub>	[46]
Dogfish	pGlu	His	Trp	Ser	His	Gly	Trp	<b>Leu</b>	Pro	Gly-NH <sub>2</sub>	[25]
Chicken I	pGlu	His	Trp	Ser	<b>Tyr</b>	Gly	<b>Leu</b>	<b>Gln</b>	Pro	Gly-NH <sub>2</sub>	[19]
Guinea pig	pGlu	<b>Tyr</b>	Trp	Ser	<b>Tyr</b>	Gly	<b>Val</b>	<b>Arg</b>	Pro	Gly-NH <sub>2</sub>	[16]
Lamprey I	pGlu	His	<b>Tyr</b>	Ser	<b>Leu</b>	<b>Glu</b>	Trp	<b>Lys</b>	Pro	Gly-NH <sub>2</sub>	[57]
Lamprey III	pGlu	His	Trp	Ser	His	<b>Asp</b>	Trp	<b>Lys</b>	Pro	Gly-NH <sub>2</sub>	[57]
Tunicate I	pGlu	His	Trp	Ser	<b>Asp</b>	Tyr	<b>Phe</b>	<b>Lys</b>	Pro	Gly-NH <sub>2</sub>	[9]
Tunicate II	pGlu	His	Trp	Ser	<b>Leu</b>	<b>Cys</b>	<b>His</b>	<b>Ala</b>	Pro	Gly-NH <sub>2</sub>	[9]

Note: The amino acids printed in bold differ from the cGnRH-II form.

### GNRH: LOCALIZATION AND ORIGIN

In general, all vertebrate species have at least two forms of GnRH (cGnRH-II and the species-specific GnRH), or even three forms, as discovered in modern fish. The two or three GnRH forms

are differentially localized in the brain (Fig. 1) and, moreover, have different embryonic origins as well.

The large cGnRH-II cells are clustered in a distinct nucleus at the fusion site of the anterior midbrain and the posterior diencephalon (the synencephalon), rostral to the oculomotor nucleus. The

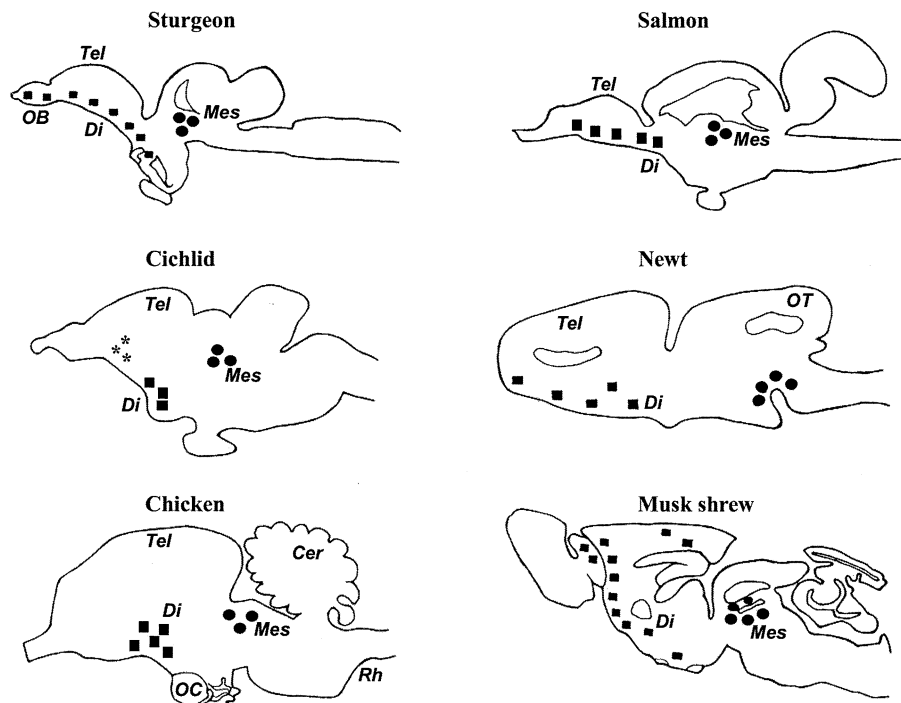


FIG. 1. Sagittal brain sections show gonadotropin-releasing hormone (GnRH) localization in sturgeon, salmon, cichlid, newt, chicken, and musk shrew. The GnRH system in the ventral forebrain is represented by ■, the cGnRH-II system in the midbrain is shown by ●, and the TN GnRH system in the cichlid is indicated by an asterisk. Images are adapted from Sherwood et al. [54]. Abbreviations: Cer, cerebellum; Di, diencephalon; Mes, mesencephalon; OB, olfactory bulb; OC, optic chiasm; OT, optic tectum; Tel, telencephalon.

TABLE 2

PHYLOGENETIC DISTRIBUTION OF GNRH FORMS ORDERED BY THEIR AREA OF LOCALIZATION IN THE BRAIN: MIDBRAIN (MB), VENTRAL FOREBRAIN (VF) OR TERMINAL NERVE (TN)

	Reference	MB	VF	TN
<b>Invertebrata</b>				
Mollusk <i>Aplysia californica</i>	[67]		m?II?	
<b>Protochordata</b>				
Tunicate <i>Ciona intestinalis</i>	[9]		m?cI? tI+II?	
<b>Agnatha</b>				
Lamprey <i>Petromyzon marinus</i>	[57]		LI+III	
Hagfish <i>Myxine glutinosa</i>	[58]		LIII	
<b>Holocephali</b>				
Ratfish <i>Hydrolagus collicie</i>	[27]	cII		
<b>Chondrichthyes</b>				
Atlantic stingray <i>Dasyatis sabina</i>	[13]	cII df	LIII?	df
Dogfish <i>Squalus acanthias</i>	[25]	cII		df
<b>Osteichthyes</b>				
<b>Reedfish <i>Calamoichthys calabaricus</i></b>				
	[51]	cII m		m
Sturgeon <i>Acipenser transmontanus</i>	[51]	cII m		m
Eel <i>Anguilla anguilla</i>	[18]	cII m		m
Herring <i>Clupea harengus pallasii</i>	[5]	cII h		s
<b>Sockeye salmon <i>Oncorhynchus nerka</i></b>				
	[42]	cII s		s
Goldfish <i>Carrasius auratus</i>	[24]	cII s		s
African catfish <i>Clarias gariepinus</i>	[3]	cII cf		cf
Pacu <i>Piaractus mesopotamicus</i>	[45]	cII sb		s
Medaka <i>Oryzias latipes</i>	[36]	cII md		s
Pejerry <i>Odentesthes bonariensis</i>	[32]	cII md		s
Red seabream <i>Pagrus major</i>	[38]	cII sb		s
Cichlid <i>Haplochromis burtoni</i>	[63]	cII sb		s
Tilapia <i>Oreochromis mossambicus</i>	[40]	cII sb		s
Lungfish <i>Protopterus annectens</i>	[22]	cII m		
<b>Amphibia</b>				
Frog <i>Rana ridibunda</i>	[6]	cII m		
Newt <i>Taricha granulosa</i>	[35]	cII m		
Frog <i>Rana dybowskii</i>	[66]	cII m	r	
<b>Reptilia</b>				
Turtle <i>Gallus domesticus</i>	[56]	cII cI		
American alligator <i>Alligator mississippiensis</i>	[26]	cII cI		
Lizard <i>Podarcis s. sicula</i>	[7]	cII cI		
<b>Aves</b>				
Chicken <i>Gallus domesticus</i>	[11]	cII cI		
Turkey <i>Gallus meleagris</i>	[30]	cII cI		
<b>Mammalia</b>				
Tree shrew <i>Tupaia glis belangeri</i>	[17]	cII m		
Guinea pig	[16]	cII gp		
Macaque <i>Macaca mulatta</i>	[23]	cII m		
Human	[62]	cII m		

In invertebrates, protochordates, and Agnathans, the localization of GnRH is unclear, which is indicated by the dashed lines between the different brain regions. Abbreviations: cf, catfish GnRH; cI, chicken GnRH-I; cII, chicken GnRH-II; df, dogfish GnRH; gp, guinea pig GnRH; h, herring GnRH; LI, lamprey GnRH-I; LIII, lamprey GnRH-III; m, mammalian GnRH; md, medaka GnRH; r, rana GnRH; s, salmon GnRH; sb, seabream GnRH; tI, tunicate GnRH-I; tII, tunicate GnRH-II.

function of this GnRH is still under debate. *In vitro* cGnRH-II can be a potent stimulator of gonadotropin release from pituitary gonadotropic cells [4], but *in vivo* levels of cGnRH-II in the

pituitary are low. The most commonly accepted function is that of neuromodulator or a role in sexual behavior [28,60]. The ventricular zone is the putative embryonic origin of these cells [41,63]. In chick embryos, this locus has been related to expression of the gene *Nkx6.1* [47].

The species-specific GnRH is localized in the ventral forebrain and mostly restricted to the ventral telencephalon, the pre-optic area (POA), the basal hypothalamus, and the pituitary gland. Because during early development neurons of this system were first observed in the olfactory bulb (OB), it has been suggested that they originate from the olfactory region. Moreover, a rare disease, Kallmann's syndrome, which is characterized by hypogonadotropic hypogonadism combined with the disability to smell, suggested an ontogenetic liaison between the olfactory system and reproductive brain areas [39,50]. It was hypothesized that the GnRH neurons of this system originate from the olfactory placode and migrate during development into the brain, colonizing the hypothalamus and the pituitary. Many studies monitored the migration of the GnRH neurons during early development in all classes of vertebrates: salmon [42], platyfish (*Xiphophorus maculatus*) [14], frog [8], lizard [7], chicken [59], rat [15], and rhesus monkey [49]. Firm evidence for this hypothesis was shown by the absence of GnRH neurons in the brain after ablation of the olfactory placode [34].

One GnRH subpopulation, the terminal nerve (TN) ganglion, is localized at the junction of the olfactory nerve and the telencephalon and is present in most vertebrates. This population has no hypophysiotropic function, but it is probably involved in reproductive behavior in relation to olfaction [65]. Its localization in close vicinity of olfactory areas suggests a neuromodulatory function in integrating olfactory signals, which may be evoked by, e.g., pheromones into the brain-pituitary axis. It is unclear whether both the TN and the POA/hypothalamus neurons are derived from the olfactory placode. Parhar et al. [40,43] hypothesized that the GnRH neurons in the terminal nerve originate from the olfactory placode, whereas GnRH neurons in the POA originate in the basal telencephalon. However, we propose that both the TN GnRH neurons and the ventral forebrain neurons originate from the olfactory placode and that they are differentially programmed concerning the timing and distance of migration.

## NOMENCLATURE

With the appearance of a third form in the brain, coexisting with cGnRH-II and the species-specific form, the terminology of the forms and systems has become complex. Therefore, Fernald and White [12] proposed a new nomenclature. GnRH1 is the species-specific form and regulates pituitary LH release; GnRH2 is the conserved cGnRH-II in the midbrain. If sGnRH is present in the terminal nerve, it is referred to as GnRH3. This nomenclature is still not widely accepted in the literature and is confusing when the GnRH form in TN and ventral forebrain is similar. Therefore, we propose an alternative nomenclature, in which both the localization and the function of the GnRH forms are combined. MB-GnRH is cGnRH-II in the midbrain; VF-GnRH is the species-specific hypophysiotropic GnRH form in the ventral forebrain; and TN-GnRH is the neuromodulatory salmon GnRH in the terminal nerve.

## EVOLUTIONARY ASPECTS OF GNRH

The presence of several molecular forms of GnRH within one species, their phylogenetic distribution, and the recent discovery of new forms [5,16,36,66] tempt one to propose a new evolutionary GnRH tree. Its construction starts with a study of the phylogenetic



4. Bosma, P. T.; Rebers, F. E. M.; Dijk, W. V.; Willems, P. H. G. M.; Goos, H. J. Th.; Schulz, R. W. Inhibitory and stimulatory interactions between endogenous gonadotropin-releasing hormones in the African catfish (*Clarias gariepinus*). *Biol. Reprod.* 62:731–738; 2000.
5. Carolsfeld, J.; Powell, J. F. F.; Park, M.; Fischer, W. H.; Craig, A. G.; Chang, J. P.; Rivier, J. E.; Sherwood, N. M. Primary structure and function of three gonadotropin-releasing hormones, including a novel form, from an ancient teleost, herring. *Endocrinology* 141:505–512; 2000.
6. Conlon, J. M.; Collin, F.; Chiang, Y. C.; Sower, S. A.; Vaudry, H. Two molecular forms of gonadotropin-releasing hormone from the brain of the frog, *Rana ridibunda*: Purification, characterization, and distribution. *Endocrinology* 132:2117–2123; 1993.
7. D'Aniello, B.; Pinelli, C.; King, J. A.; Rastogi, R. K. Neuroanatomical organization of GnRH neuronal systems in the lizard (*Podarcis sicula*) brain during development. *Brain Res.* 657:221–226; 1994.
8. Di Fiore, M. M.; King, J. A.; D'Aniello, B.; Rastogi, R. K. Immunoreactive mammalian and chicken-II GnRHs in *Rana esculenta* brain during development. *Reg. Peptides* 62:119–124; 1996.
9. Di Fiore, M. M.; Rastogi, R. K.; Ceciliani, F.; Messi, E.; Botte, V.; Botte, L.; Pinelli, C.; D'Aniello, B.; D'Aniello, A. Mammalian and chicken I forms of gonadotropin-releasing hormone in the gonads of a protochordate, *Ciona intestinalis*. *Proc. Natl. Acad. Sci. USA* 97:2343–2348; 2000.
10. Dubois, E. A. Development of gonadotropin-releasing hormone systems in the male African catfish, *Clarias gariepinus*. PhD Thesis. ISBN 90-393-2645-2:13–48; 2001.
11. Dunn, I. C.; Millam, J. R. Gonadotropin releasing hormone: Forms and functions in birds. *Poultry Avian Biol. Rev.* 9:61–85; 1998.
12. Fernald, R. D.; White, R. B. Gonadotropin-releasing hormone genes: Phylogeny, structure, and functions. *Front. Neuroendocrin.* 20:224–240; 1999.
13. Forlano, P. M.; Maruska, K. P.; Sower, S. A.; King, J. A.; Tricas, T. C. Differential distribution of gonadotropin-releasing hormone-immunoreactive neurons in the stingray brain: Functional and evolutionary considerations. *Gen. Comp. Endocrinol.* 118:226–248; 2000.
14. Halpern-Sebold, L. R.; Schreibman, M. P. Ontogeny of centers containing luteinizing hormone-releasing hormone in the brain of platyfish (*Xiphophorus maculatus*) as determined by immunocytochemistry. *Cell Tissue Res.* 229:75–84; 1983.
15. Jennes, L. Prenatal development of the gonadotropin-releasing hormone-containing systems in the brain. *Brain Res.* 482:97–108; 1989.
16. Jimenez-Linan, M.; Rubin, B. S.; King, J. C. Examination of guinea pig luteinizing hormone-releasing hormone gene reveals a unique decapeptide and existence of two transcripts in the brain. *Endocrinology* 138:4123–4130; 1997.
17. Kasten, T. L.; White, S. A.; Norton, T. T.; Bond, C. T.; Adelman, J. P.; Fernald, R. D. Characterization of two new preproGnRH mRNAs in the tree shrew: First direct evidence for mesencephalic GnRH gene expression in a placental mammal. *Gen. Comp. Endocrinol.* 104:7–19; 1996.
18. King, J. A.; Dufour, S.; Fontaine, Y. A.; Millar, R. P. Chromatographic and immunological evidence for mammalian GnRH and chicken GnRH-II in eel (*Anguilla anguilla*) brain and pituitary. *Peptides* 11:507–514; 1990.
19. King, J. A.; Millar, R. P. Structure of chicken hypothalamic luteinizing hormone-releasing hormone. II. Isolation and characterization. *J. Biol. Chem.* 257:10729–10732; 1982.
20. King, J. A.; Millar, R. P. Evolution of gonadotropin-releasing hormones. *TEM* 3:339–346; 1992.
21. King, J. A.; Millar, R. P. Coordinated evolution of GnRHs and their receptors. In: Parhar, I. S.; Sakuma, Y., eds. *GnRH neurons: Gene to behavior*. Tokyo: Brain Shuppan; 1998:51–77.
22. King, J. A.; Millar, R. P.; Vallarino, M.; Pierantoni, R. Localization and characterization of gonadotropin-releasing hormones in the brain, gonads, and plasma of a dipnoi (lungfish, *Protopterus annectens*). *Reg. Peptides* 57:163–174; 1995.
23. Latimer, V. S.; Rodrigues, S. M.; Garyfallou, V. T.; Kohama, S. G.; White, R. B.; Fernald, R. D.; Urbanski, H. F. Two molecular forms of gonadotropin-releasing hormone (GnRH-I and GnRH-II) are expressed by two separate populations of cells in the rhesus macaque hypothalamus. *Mol. Brain Res.* 75:287–292; 2000.
24. Lin, X. W.; Peter, R. E. Cloning and expression pattern of a second [His<sup>5</sup>Trp<sup>7</sup>Tyr<sup>8</sup>]gonadotropin-releasing hormone (chickenGnRH-II) mRNA in goldfish: Evidence for two distinct genes. *Gen. Comp. Endocrinol.* 170:262–272; 1997.
25. Lovejoy, D. A.; Fischer, W. H.; Ngamvongchon, S.; Craig, A. G.; Nahorniak, C. S.; Peter, R. E.; Rivier, J. E.; Sherwood, N. M. Distinct sequence of gonadotropin-releasing hormone (GnRH) in dogfish brain provides insight into GnRH evolution. *Proc. Natl. Acad. Sci. USA* 89:6373–6377; 1992.
26. Lovejoy, D. A.; Fischer, W. H.; Parker, D. B.; McRory, J. E.; Park, M.; Lance, V.; Swanson, P.; Rivier, J. E.; Sherwood, N. M. Primary structure of two forms of gonadotropin-releasing hormone from brains of the American alligator (*Alligator mississippiensis*). *Reg. Peptides* 33:105–116; 1991.
27. Lovejoy, D. A.; Sherwood, N. M.; Fischer, W. H.; Jackson, B. C.; Rivier, J. E.; Lee, T. Primary structure of gonadotropin-releasing hormone from the brain of a holocephalan (ratfish: *Hydrolagus coliei*). *Gen. Comp. Endocrinol.* 82:152–161; 1991.
28. Maney, D. L.; Richardson, R. D.; Wingfield, J. C. Central administration of chicken gonadotropin-releasing hormone-II enhances courtship behavior in a female sparrow. *Horm. Behav.* 32:11–18; 1997.
29. Matsuo, H.; Baba, Y.; Nair, N. M. G.; Arimura, A.; Schally, A. V. Structure of the porcine LH- and FSH-releasing hormone. I. The proposed amino acid sequence. *Biochem. Biophys. Res. Commun.* 43:1334–1339; 1971.
30. Millam, J. R.; Faris, P. L.; Youngren, O. M.; El Halawani, M. E.; Hartman, B. K. Immunohistochemical localization of chicken gonadotropin-releasing hormones I and II (cGnRH I and II) in turkey hen brain. *J. Comp. Neurol.* 333:68–82; 1993.
31. Miyamoto, K.; Hasegawa, Y.; Nomura, M.; Igarashi, M.; Kangawa, K.; Matsuo, H. Identification of the second gonadotropin-releasing hormone in chicken hypothalamus: Evidence that gonadotropin secretion is probably controlled by two distinct gonadotropin-releasing hormones in avian species. *Proc. Natl. Acad. Sci. USA* 81:3874–3878; 1984.
32. Montaner, A. D.; Park, M. K.; Fischer, W. H.; Craig, A. G.; Chang, J. P.; Somoza, G. M.; Rivier, J. E.; Sherwood, N. M. Primary structure of a novel gonadotropin-releasing hormone in the brain of a teleost, Pejerrey. *Endocrinology* 142:1453–1460; 2001.
33. Montero, M.; Dufour, S. Gonadotropin-releasing hormones (GnRH) in fishes: Evolutionary data on their structure, localization, regulation, and function. *Zool. Stud.* 35:149–160; 1996.
34. Murakami, S.; Kikuyama, S.; Arai, Y. The origin of the luteinizing hormone-releasing hormone (LHRH) neurons in newts (*Cynops pyrrhogaster*): The effect of olfactory placode ablation. *Cell Tissue Res.* 269:21–27; 1992.
35. Muske, L. E.; Moore, F. L. Antibodies against different forms of GnRH distinguish different populations of cells and axonal pathways in a urodele amphibian, *Taricha granulosa*. *J. Comp. Neurol.* 345:139–147; 1994.
36. Okubo, K.; Amano, M.; Yoshiura, Y.; Suetake, H.; Aida, K. A novel form of gonadotropin-releasing hormone in the medaka, *Oryzias latipes*. *Biochem. Biophys. Res. Commun.* 276:298–303; 2000.
37. Okubo, K.; Suetake, H.; Aida, K. Expression of two gonadotropin-releasing hormone (GnRH) precursor genes in various tissues of the Japanese eel and evolution of GnRH. *Zool. Sci.* 16:471–478; 1999.
38. Okuzawa, K.; Granneman, J.; Bogerd, J.; Goos, H. J. Th.; Zohar, Y.; Kagawa, H. Distinct expression of GnRH genes in the red seabream. *Fish Physiol. Biochem.* 17:71–79; 1997.
39. Parhar, I.; Pfaff, D.; Schwanzel-Fukuda, M. Genes and behavior as studied through gonadotropin-releasing hormone (GnRH) neurons: Comparative and functional aspects. *Cell. Mol. Neurobiol.* 15:107–116; 1995.
40. Parhar, I. S. GnRH in Tilapia: Three genes, three origins and their roles. In: Parhar, I. S.; Sakuma, Y., eds. *GnRH neurons, gene to behavior*. Tokyo: Brain Shuppan; 1998:99–122.
41. Parhar, I. S. Multiple gonadotropin-releasing hormone neuronal systems in vertebrates. *Korean J. Biol. Sci.* 3:1–7; 1999.
42. Parhar, I. S.; Iwata, M.; Pfaff, D. W.; Schwanzel-Fukuda, M. Embryonic development of gonadotropin-releasing hormone neurons in the sockeye salmon. *J. Comp. Neurol.* 362:256–270; 1995.
43. Parhar, I. S.; Soga, T.; Ishikawa, Y.; Nagahama, Y.; Sakuma, Y.

- Neurons synthesizing gonadotropin-releasing hormone mRNA subtypes have multiple developmental origins in the medaka. *J. Comp. Neurol.* 401:217–226; 1998.
44. Powell, J. F. F.; Reskaskinner, S. M.; Prakash, M. O.; Fischer, W. H.; Park, M.; Rivier, J. E.; Craig, A. G.; Mackie, G. O.; Sherwood, N. M. Two new forms of gonadotropin-releasing hormone in a protochordate and the evolutionary implications. *Proc. Natl. Acad. Sci. USA* 93:10461–10464; 1996.
  45. Powell, J. F. F.; Standen, E. M.; Carolsfeld, J.; Borella, M. I.; Gazola, R.; Fischer, W. H.; Park, M.; Craig, A. G.; Warby, C. M.; Rivier, J. E.; Val-Sella, M. V.; Sherwood, N. M. Primary structure of three forms of gonadotropin-releasing hormone (GnRH) from the pacu brain. *Reg. Peptides* 68:189–195; 1997.
  46. Powell, J. F. F.; Zohar, Y.; Elizur, A.; Park, M.; Fischer, W. H.; Craig, A. G.; Rivier, J. E.; Lovejoy, D. A.; Sherwood, N. M. Three forms of gonadotropin-releasing hormone characterized from brains of one species. *Proc. Natl. Acad. Sci. USA* 91:12081–12085; 1994.
  47. Puelles, E.; Rubenstein J. L.; Puelles, L. Chicken Nkx6.1 expression at advanced stages of development identifies distinct brain nuclei derived from the basal plate. *Mech. Dev.* 102:279–282; 2001.
  48. Quanbeck, C.; Sherwood, N. M.; Millar, R. P.; Terasawa, E. Two populations of luteinizing hormone-releasing hormone neurons in the forebrain of the rhesus macaque during embryonic development. *J. Comp. Neurol.* 380:293–309; 1997.
  49. Ronnekleiv, O. K.; Resko, J. A. Ontogeny of gonadotropin-releasing hormone-containing neurons in early fetal development of rhesus macaques. *Endocrinology* 126:498–511; 1990.
  50. Schwanzel-Fukuda, M.; Jorgenson, K. L.; Bergen, H. T.; Weesner, G. D.; Pfaff, D. W. Biology of normal luteinizing hormone-releasing hormone neurons during and after their migration from olfactory placode. *Endocrine Rev.* 13:623–634; 1992.
  51. Sherwood, N. M.; Doroshov, S.; Lance, V. Gonadotropin-releasing hormone (GnRH) in bony fish that are phylogenetically ancient: Reedfish (*Calamoichthys calabaricus*), sturgeon (*Acipenser transmontanus*), and alligator gar (*Lepisosteus spatula*). *Gen. Comp. Endocrinol.* 84:44–57; 1991.
  52. Sherwood, N. M.; Eiden, L.; Rivier, J. E.; Spiess, J.; Vale, W. Characterization of a teleost gonadotropin-releasing hormone. *Proc. Natl. Acad. Sci. USA* 80:2794–2798; 1983.
  53. Sherwood, N. M.; Lovejoy, D. A.; Coe, I. R. Origin of mammalian gonadotropin-releasing hormones. *Endocrine Rev.* 14:241–254; 1993.
  54. Sherwood, N. M.; Schalburg, K. R. v.; Lescheid, D. W. Origin and evolution of GnRH in vertebrates and invertebrates. In: Parhar, I. S.; Sakuma, Y., eds. *GnRH neurons: Gene to behavior*. Tokyo: Brain Shuppan; 1997:3–25.
  55. Sherwood, N. M.; Sower, S. A.; Marshak, D. R.; Fraser, B. A.; Brownstein, M. J. Primary structure of gonadotropin-releasing hormone from lamprey brain. *J. Biol. Chem.* 261:4812–4819; 1986.
  56. Sherwood, N. M.; Whittier, J. M. Gonadotropin-releasing hormone from brains of reptiles: Turtles (*Pseudemys scripta*) and snakes (*Thamnophis sirtalis parietalis*). *Gen. Comp. Endocrinol.* 69:319–327; 1988.
  57. Sower, S. A.; Chiang, Y. C.; Lovas, S.; Conlon, J. M. Primary structure and biological activity of a third gonadotropin-releasing hormone from lamprey brain. *Endocrinology* 132:1125–1131; 1993.
  58. Sower, S. A.; Nozaki, M.; Knox, C. J.; Gorbman, A. The occurrence and distribution of GnRH in the brain of Atlantic hagfish, an agnathan, determined by chromatography and immunocytochemistry. *Gen. Comp. Endocrinol.* 97:300–307; 1995.
  59. Sullivan, K. A.; Silverman, A. J. The ontogeny of gonadotropin-releasing hormone neurons in the chick. *Neuroendocrinology* 58:597–608; 1993.
  60. Volkoff, H.; Peter, R. E. Actions of two forms of gonadotropin-releasing hormone and a GnRH antagonist on spawning behavior of the goldfish *Carrasius auratus*. *Gen. Comp. Endocrinol.* 116:347–355; 1999.
  61. Wang, L.; Yoo, M. S.; Kang, H. M.; Im, W. B.; Choi, H. S.; Bogerd, J.; Kwon, H. B. Cloning and characterization of cDNAs encoding the GnRH1 and GnRH2 precursors from bullfrog (*Rana catesbeiana*). *J. Exp. Zool.* 289:190–201; 2001.
  62. White, R. B.; Eisen, J. A.; Kasten, T. L.; Fernald, R. D. Second gene for gonadotropin-releasing hormone in humans. *Proc. Natl. Acad. Sci. USA* 95:305–309; 1998.
  63. White, R. B.; Fernald, R. D. Ontogeny of gonadotropin-releasing hormone (GnRH) gene expression reveals a distinct origin for GnRH-containing neurons in the midbrain. *Gen. Comp. Endocrinol.* 112:322–329; 1998.
  64. White, S. A.; Kasten, T. L.; Bond, C. T.; Adelman, J. P.; Fernald, R. D. Three gonadotropin-releasing hormone genes in one organism suggest novel roles for an ancient peptide. *Proc. Natl. Acad. Sci. USA* 92:8363–8367; 1995.
  65. Yamamoto, N.; Oka, Y.; Kawashima, S. Lesions of gonadotropin-releasing hormone-immunoreactive terminal nerve cells: Effects on the reproductive behavior of male dwarf gouramis. *Neuroendocrinology* 65:403–412; 1997.
  66. Yoo, M. S.; Kang, H. M.; Choi, H. S.; Kim, J. W.; Troskie, B. E.; Millar, R. P.; Kwon, H. B. Molecular cloning, distribution and pharmacological characterization of a novel gonadotropin-releasing hormone ([Trp8]GnRH) in frog brain. *Mol. Cell. Endocrinol.* 164:197–204; 2000.
  67. Zhang, L.; Wayne, N. L.; Sherwood, N. M.; Postigo, H. R.; Tsai, P. S. Biological and immunological characterization of multiple GnRH in an opisthobranch mollusk, *Aplysia californica*. *Gen. Comp. Endocrinol.* 117:77–89; 2000.